AMENDMENTS TO THE SPECIFICATION

<u>Please replace the paragraph on page 1, lines 2-9 with the following amended paragraph:</u>

This application is a continuation application of U.S.

Serial No. 08/260,190 (filed June 15, 1994) which is a

continuation-in-part of now pending U.S. Serial No. 08/177,093

(filed December 30, 1993) which issued on April 18, 2000 as U.S.

Patent No. 6,051,226, which is in turn a continuation-in-part of

U.S. Serial No. 07/964,589 (filed October 21, 1992) which issued

on February 7, 1995 as U.S. Patent No. 5,387,676. This

application declares priority under 35 USC § 120 from those U.S.

applications, and also under 35 USC § 119 from the now pending

Czechoslovakian patent application PV-709-92 (filed March 11, 1992).

Please replace the paragraph on page 8, lines 3-11 with the following amended paragraph:

This invention also concerns nucleic acids which encode MN proteins or polypeptides that are specifically bound by monoclonal antibodies designated M75 that are produced by the hybridoma VU-M75 deposited at the American Type Culture Collection (ATCC) at 12301 Parklawn Drive in Rockville, Maryland 20852 10801 University Blvd., Manassas, Virginia 20110-2209 (USA) under ATCC No. HB 11128, and/or by monoclonal antibodies designated MN12 produced by the hybridoma MN 12.2.2 deposited at the ATCC under ATCC No. HB 11647.

Please replace the paragraph on page 13, lines 3-14 with the following amended paragraph:

A hybridoma that produces a representative MN-specific antibody, the monoclonal antibody M75 (Mab M75), was deposited at the under ATCC atcc under Number HB 11128 as indicated above.

The M75 antibody was used to discover and identify the MN protein and can be used to identify readily MN antigen in Western blots, in radioimmunoassays and immunohistochemically, for example, in tissue samples that are fresh, frozen, or formalin-, alcohol-, acetone- or otherwise fixed and/or paraffin-embedded and deparaffinized. Another representative MN-specific antibody, Mab MN12, is secreted by the hybridoma MN 12.2.2, which was deposited at the ATCC under the designation HB 11647.

Please replace the paragraph on page 16, lines 3-10 with the following amended paragraph:

The immunoassays of this invention can be embodied in test kits which comprise MN proteins/polypeptides and/or MN-specific antibodies. Such test kits can be in solid phase formats, but are not limited thereto, and can also be in liquid phase format, and can be based on immunohistochemical assays, ELISAS, particle assays, radiometric or fluorometric assays either unamplified or amplified, using, for example, avidin/biotin technology.

<u>Please replace line 26 on page 17 with the following amended line:</u>

IPTG - <u>isopropyl-Beta</u> <u>isopropyl-beta</u>-D-thiogalactopyranoside

<u>Please replace the paragraph bridges pages 27 and 28 with the following amended paragraph:</u>

Figure 15 Figure 15A-C shows a complete nucleotide sequence of a MN cDNA [SEQ. ID. NO.: 5]. Also shown is the deduced amino acid sequence [SEQ. ID. NO.: 6]. The polyadenylation signal (AATAAA) and the mRNA instability motif (ATTTA) are underlined are located at nucleotides (nts) 1507-1512 and at nts 1513-1518, respectively. The amino acid residues of the putative signal peptide as well as the membrane-spanning segment are italicized are located at amino acids (aa) 1-37 and at aa 415-433, respectively. The N-glycosylation site and the putative nuclear localization signal are denoted by squares and asterisks, respectively is located at aa 346. The S/TPXX elements are indicated with open circles are located at amino acids 7-10, 47-50, 71-74, 153-156, 162-165, 333-336, and 397-400.

<u>Please replace the paragraph on page 54, lines 14-20 with the following amended paragraph:</u>

Where the host used is an eucaryote, eukaryote, transfection methods such as the use of a calcium phosphate-precipitate, electroporation, conventional mechanical procedures such as microinjection, insertion of a plasmid encapsulated in red blood cell ghosts or in liposomes, treatment of cells with

agents such as lysophosphatidyl-choline or use of virus vectors, or the like may be used.

<u>Please replace the paragraph on page 54, lines 14-20 with the following amended paragraph:</u>

The MN 20-19 protein was purified from the conditioned media by immunoaffinity chromatography. 6.5 mg of Mab M75 was coupled to 1 g of Tresyl activated Toyopearl™ [solid support in bead form; Tosoh, Japan (#14471)]. Approximately 150 ml of the conditioned media was run through the M75-Toyopearl column. The column was washed with PBS, and the MN 20-19 protein was eluted with 1.5 M MgCl. The eluted protein was then dialyzed against PBS.

<u>Please replace the paragraph on page 86, lines 10-16 with the following amended paragraph:</u>

MAD M75. Monoclonal antibody M75 (MAD M75) is produced by mouse lymphocytic hybridoma VU-M75, which was initially deposited in the Collection of Hybridomas at the Institute of Virology, Slovak Academy of Sciences (Bratislava, Czechoslovakia Slovakia) and was deposited under ATCC Designation HB 11128 on September 17, 1992 at the American Type Culture Collection (ATCC) in Rockville, MD Manassas, Virginia (USA).

Please replace the paragraph on page 89, lines 9-20 with the following amended paragraph:

Mab MN12. Monoclonal antibody MN12 (Mab MN12) is produced by the mouse lymphocytic hybridoma MN 12.2.2 which was deposited under ATCC Designation HB 11647 on June 9, 1994 at the American Type Culture Collection (ATCC) at 12301 Parklawn Drive, Rockville, MD 20852 10801 University Blvd., Manassas, Virginia 20110-2209 (USA). Antibodies corresponding to Mab MN12 can also be made, analogously to the method outlined above for Mab MN9, by screening a series of antibodies prepared against an MN protein/polypeptide, against the peptide representing the epitope for Mab MN12. That peptide is Gly Lys Met Thr His Trp [SEQ. ID. NO.: 11]. The Novatope system could also be used to find antibodies specific for said epitope.

Please delete line 23 on page 91:

Antisense MN Nucleic Acid Sequences

<u>Please replace the paragraph bridges pages 94 and 95 with the</u> following amended paragraph:

MN proteins and/or polypeptides may be synthesized or prepared recombinantly or otherwise biologically, to comprise one or more amino acid sequences corresponding to one or more epitopes of the MN proteins either in monomeric or multimeric form. Those proteins and/or polypeptides may then be incorporated into vaccines capable of inducing protective immunity. Techniques for enhancing the antigenicity of such

polypeptides include incorporation into a multimeric structure, binding to a highly immunogenic protein carrier, for example, keyhole limpet hemocyanin (KLH), or diptheria diphtheria toxoid, and administration in combination with adjuvants or any other enhancers of immune response.

Please replace the paragraph bridges pages 95 and 96 with the following amended paragraph:

An amino acid sequence corresponding to an epitope of an MN protein/polypeptide either in monomeric or multimeric form may also be obtained by chemical synthetic means or by purification from biological sources including genetically modified microorganisms or their culture media. [See Lerner, "Synthetic Vaccines", <u>Sci. Am. 248(2)</u>: 66-74 (1983).] protein/polypeptide may be combined in an amino acid sequence with other proteins/polypeptides including fragments of other proteins, as for example, when synthesized as a fusion protein, or linked to other antigenic or non-antigeneic non-antigenic polypeptides of synthetic or biological origin. instances, it may be desirable to fuse a MN protein or polypeptide to an immunogenic and/or antigenic protein or polypeptide, for example, to stimulate efficacy of a MN-based vaccine.

Please replace the two paragraphs on page 98, lines 11-24 with the following amended paragraphs:

Human sera from cancer patients, from patients suffering with various non-tumor complaints and from healthy

women were obtained from the Clinics of Obstetrics and Gynaecology at the Postgraduate Medical School, Bratislava, Czechoslovakia Slovakia. Human sera serum KH was from a fifty year old mammary carcinoma patient, fourteen months after resection. That serum was one of two sera out of 401 serum samples that contained neutralizing antibodies to the VSV(MaTU) pseudotype as described in Zavada et al. (1972), supra. Serum L8 was from a patient with Paget's disease. Serum M7 was from a healthy donor.

Rabbit anti-MaTu serum was prepared by immunizing a rabbit three times at intervals of 30 days with 10-5 days with 1-5 x 10^7 viable MaTu-infected HeLa cells.

<u>Please replace the paragraph on page 107, lines 4-11 with the following amended paragraph:</u>

A radimmunoassay radioimmunoassay was performed directly in confluent petri dish (5 cm) culture of cells, fixed with methanol essentially as described in Example 3, supra. The monolayers were fixed with methanol and treated with 125I-labeled MAbs M67 (specific for exogenous MX antigen) or M75 (specific for endogenous MN antigen) at 6 x 104 cpm/dish. The bound radioactivity was measured; the results are shown in Figure 6.

<u>Please replace the paragraph bridging pages 119 and 120 with the following amended paragraph:</u>

It was found that cultivation of HeLa cells with the ODNs resulted in considerable inhibition of p54/58N synthesis.

The 19-mer ODN2 (Figure 3B) in 4 μ M final concentration was very effective; as determined by RIA, it caused 40% inhibition, whereas the 29-mer ODN1 (4 μ M) (Figure 3A) and a combination of the two ODNs (Figure 3C), each in 2 μ M final concentration, were less effective in RIA showing a 25-35% increase decrease of the MN-related proteins. At the same time, the amount of different HeLa cell protein determined by RIA using specific MAb H460 was in all cell variants approximately the same. Most importantly was that on immunoblot it could be seen that specific inhibition by the ODNs affected both of the p54/58N proteins. Thus, we concluded that the MN gene we cloned coded for both p54/58N proteins in HeLa cells.

Please replace TABLE 2 on page 131, lines 1-27 with the following amended TABLE 2:

TABLE 2

Immunoreactivity of M75 in Various Tissues

TISSUE	TYPE	POS/NEG (#pos/#tested)					
liver, spleen, lung, kidney, adrenal gland, brain, prostate, pancreas, thyroid,							
<pre>ovary, testis</pre>	normal	NEG (all)					
skin	normal	POS (in basal layer) (1/1)					
stomach	normal	POS					
small intestine	normal	POS					
colon	normal	POS					
breast	normal	NEG (0/10)					
cervix	normal	NEG (0/2)					
breast	benign	NEG (0/17)					
colon	benign	POS (4/11)					
cervix	benign	POS (10/18)					
breast	malignant	POS (3/25)					
colon	malignant	POS (9/15)					
ovarian	malignant	POS (3/15)					
lung	malignant	POS (12/30)					
bladder head & neck	malignant malignant	POS (1/3) POS (3/4)					
kidney	malignant	POS (3/4) POS (4/4)					
stomach	malignant	NEG (0/4)					
cervix	malignant	POS (62/68)					
	<u>J</u>	·,					

Please replace the paragraph bridging pages 141 and 142 with the following amended paragraph:

ATCC Deposits. The material listed below was deposited with the American Type Culture Collection (ATCC) at 12301

Parklawn Drive, Rockville, MD 20852 10801 University Blvd.,

Manassas, Virginia 20110-2209 (USA). The deposits were made

under the provisions of the Budapest Treaty on the International Recognition of Deposited Microorganisms for the Purposes of Patent Procedure and Regulations thereunder (Budapest Treaty). Maintenance of a viable culture is assured for thirty years from the date of deposit. The organism will be made available by the ATCC under the terms of the Budapest Treaty, and subject to an agreement between the Applicants and the ATCC which assures unrestricted availability of the deposited hybridomas to the public upon the granting of patent from the instant application. Availability of the deposited strain is not to be construed as a license to practice the invention in contravention of the rights granted under the authority of any Government in accordance with its patent laws.

Page 143 After the Detailed Description, please insert the SEQUENCE LISTING:

SEQUENCE LISTING

<110> Zavada, Jan
 Pastorekova, Silvia
 Pastorek, Jaromir

<120> MN Gene and Protein

<130> D-0021.2-2

<140>

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<150> 08/260,190

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gag gag gat cta cct gga gag gat cta cct gaa gtt aag cct aaa Glu Glu Asp Leu Pro Gly Glu Glu Asp Leu Pro Glu Val Lys Pro Lys 50 55

144

192

tca Ser 65	gaa Glu	gaa Glu	gag Glu	ggc Gly	tcc Ser 70	ctg Leu	aag Lys	tta Leu	gag Glu	gat Asp 75	cta Leu	cct Pro	act Thr	gtt Val	gag Glu 80	240
gct Ala	cct Pro	gga Gly	gat Asp	cct Pro 85	caa Gln	gaa Glu	ccc Pro	cag Gln	aat Asn 90	aat Asn	gcc Ala	cac His	agg Arg	gac Asp 95	aaa Lys	288
gaa Glu	GIY ggg	gat Asp	gac Asp 100	cag Gln	agt Ser	cat His	tgg Trp	cgc Arg 105	tat Tyr	gga GIy	ggc	gac Asp	ccg Pro 110	ccc Pro	tgg Trp	336
ccc Pro	cgg Arg	gtg Val 115	tcc Ser	cca Pro	gcc Ala	tgc Cys	gcg Ala 120	ggc Gly	cgc Arg	ttc Phe	cag Gln	tcc Ser 125	ccg Pro	gtg Val	gat Asp	384
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ctc Leu 145	ctg Leu	ggc	ttc Phe	cag Gln	ctc Leu 150	ccg Pro	ccg Pro	ctc Leu	cca Pro	gaa Glu 155	ctg Leu	cgc Arg	ctg Leu	cgc Arg	aac Asn 160	480
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					_	act Thr 295		_		_	_	_		_		912
tgg Trp 305	act Thr	gtg Val	ttt Phe	aac Asn	cag Gln 310	aca Thr	gtg Val	atg Met	ctg Leu	agt Ser 315	gct Ala	aag Lys	cag Gln	ctc Leu	cac His 320	960
acc Thr	ctc Leu	tct Ser	gac Asp	acc Thr 325	ctg Leu	tgg Trp	gga Gly	cct Pro	ggt Gly 330	gac Asp	tct Ser	cgg Arg	cta Leu	cag Gln 335	ctg Leu	1008

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Let cct gct gga gtg gac agc ser ser pro Arg Ala Ala Glu pro Val Gln

Ctg aat tcc tgc ctg gct gct ggt gac atc cta gcc ctg gtt ttt ggc

Leu Asn Ser Cys Leu Ala Ala Gly Asp Ile Leu Ala Leu Val Phe Gly

370

Ctc ctt ttt gct gtc acc agc gtc gcg ttc ctt gtg agc atc cta gcc ctg gtt ttt ggc

Leu Phe Ala Val Thr Ser Val Ala Phe Leu Val Gln Met Arg Arg

385

Ctc aga acc aga agg gga acc acc ggg gt gac atc cta gcc ctg gtt ttt ggc

Leu Asn Ser Cys Leu Ala Ala Phe Leu Ala Leu Val Phe Gly

385

Ctc ctt ttt gct gct acc aga ggg ggt gt gtg agc tac cgc ctg gt acc aga atg

Leu Asn Arg Arg Gly Thr Lys Gly Gly Val Ser Tyr Arg Pro Ala Glu

415

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1296

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